HUMAN SOLUBLE uPAR ELISA

Catalogue number: **REH001R**

Instructions for Use:

For research use only!





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HISTORY OF CHANGES

Previous version	Current version
	ENG.001.A
New edition.	

1. INTRODUCTION

BioVendor Human soluble uPAR (urokinase-type plasminogen activator receptor, uPAR: suPAR, soluble plasminogen activator urokinase receptor, soluble urokinase-type plasminogen activator receptor, PLAUR, sPLAUR) ELISA kit is an in vitro enzyme-linked immunosorbent assay for the quantitative measurement of human suPAR in serum, plasma, and cell culture supernatants. The assay employs an antibody specific for human suPAR coated on a 96-well plate. Standards and samples are pipetted into the wells and suPAR present in a sample is bound to the wells by the immobilized antibody. The wells are washed and biotinylated anti-human suPAR antibody is added. After washing away unbound biotinylated antibody, HRP-conjugated streptavidin is pipetted to the wells. The wells are again washed, a TMB substrate solution is added to the wells and color develops in proportion to the amount of suPAR bound. The Stop Solution changes the color from blue to yellow, and the intensity of the color is measured at 450 nm.

2. STORAGE, EXPIRATION

The kit may be stored at 4°C for up to 6 months. For extended storage, it is recommended to store at -80°C. For prepared reagent storage, see table below.

3. REAGENT SUPPLIED

Component	Size / Description	Storage / Stability After Preparation
suPAR Microplate (Item A)	96 wells (12 strips x 8 wells) coated with anti-Human suPAR	1 month at 4 °C
Wash Buffer Concentrate (20X) (Item B)	25 ml of 20X concentrated solution	1 month at 4 °C
Standard Protein (Item C)	2 vials of Human suPAR. 1 vial is enough to run each standard in duplicate	1 week at -80 °C
Detection Antibody suPAR (Item F)	2 vials of biotinylated anti-Human suPAR. Each vial is enough to assay half the microplate	5 days at 4 °C
HRP-Streptavidin Concentrate (Item G)	200 µl 400X concentrated HRP- conjugated strepavidin	Do not store and reuse
TMB One-Step Substrate Reagent (Item H)	12 ml of 3,3,5,5'-tetramethylbenzidine (TMB) in buffer solution	N/A
Stop Solution (Item I)	8 ml of 0.2 M sulfuric acid	N/A
Assay Diluent (Item E2)	15ml of 5X concentrated buffer	1 month at 4 °C

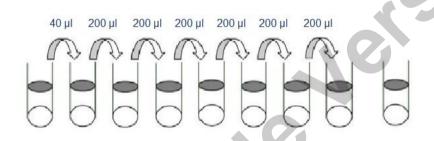
Return unused wells to the pouch containing desiccant pack, reseal along entire edge!

4. MATERIAL REQUIRED BUT NOT SUPPLIED

- 1. Microplate reader capable of measuring absorbance at 450 nm
- 2. Precision pipettes to deliver 2 µl to 1 ml volume
- 3. Adjustable 1-25 ml pipettes for reagent preparation
- 4. 100 ml and 1-liter graduated cylinders
- 5. Absorbent paper
- 6. Distilled or deionized water
- 7. Log-log graph paper or computer and software for ELISA data analysis
- 8. Tubes to prepare standard or sample dilution

5. PREPARATION OF REAGENTS

- 1. Bring all reagents and samples to room temperature (18 25°C) before use.
- 2. Assay Diluent (Item E2) should be diluted 5-fold with deionized or distilled water before use.
- 3. Sample dilution: 1X Assay Diluent (Item E2) should be used for dilution of serum, plasma, and cell culture supernatant samples. The suggested dilution for normal serum/plasma is 2-10 fold.
 - <u>Note:</u> Levels of suPAR may vary between different samples. Optimal dilution factors for each sample must be determined by the investigator.
- 4. Preparation of standard: Briefly spin a vial of Item C. Add 400 μl 1X Assay Diluent (Item E2; Assay Diluent should be diluted 5-fold with deionized or distilled water before use) into Item C vial to prepare a 50 ng/ml standard solution. Dissolve the powder thoroughly by a gentle mix. Add 40 μl of the suPAR standard from the vial of Item C, into a tube with 460 μl 1X Assay Diluent to prepare a 4 000 pg/ml standard solution. Pipette 300 μl 1X Assay Diluent into each tube. Use the 4 000 pg/ml standard solution to produce a dilution series (shown below). Mix each tube thoroughly before the next transfer. 1X Assay Diluent serves as the zero standard (0 pg/ml).



		Std1	Std2	Std3	Std4	Std5	Std6	Std7	Zero Standard
Diluent volume	Item C+400 µI	460 µl	300 µl	300 µl	300 µl	300 µl	300 µl	300 µl	300 µl
Conc.	50 ng/ml	4000 pg/ml	1600 pg/ml	640 pg/ml	256 pg/ml	102.4 pg/ml	40.96 pg/ml	16.38 pg/ml	0 pg/ml

- 5. If the Wash Concentrate (20X) (Item B) contains visible crystals, warm to room temperature and mix gently until dissolved. Dilute 20 ml of Wash Buffer Concentrate into deionized or distilled water to yield 400 ml of 1X Wash Buffer.
- 6. Briefly spin the Detection Antibody vial (Item F) before use. Add 100 µl of 1X Assay Diluent (Item E2) into the vial to prepare a detection antibody concentrate. Pipette up and down to mix gently (the concentrate can be stored at 4°C for 5 days). The detection antibody concentrate should be diluted 80-fold with 1X Assay Diluent (Item E2) and used in step 5 of Part VI Assay Procedure.
- 7. Briefly spin the HRP-Streptavidin concentrate vial (Item G) and pipette up and down to mix gently before use, as precipitates may form during storage. HRP-Streptavidin concentrate should be diluted 400-fold with 1X Assay Diluent (Item E2).

For example: Briefly spin the vial (Item G) and pipette up and down to mix gently. Add 30 µl of HRP-Streptavidin concentrate into a tube with 12 ml 1X Assay Diluent to prepare a 400-fold diluted HRP-Streptavidin solution (don't store the diluted solution for next day use). Mix well.

6. ASSAY PROCEDURE

- 1. Bring all reagents and samples to room temperature (18 25°C) before use. It is recommended that all standards and samples be run at least in duplicate.
- 2. Label removable 8-well strips as appropriate for your experiment.
- 3. Add 100 µl of each standard (see Reagent Preparation step 3) and sample into appropriate wells. Cover wells and incubate for 2.5 hours at room temperature with gentle shaking.
- 4. Discard the solution and wash 4 times with 1X Wash Solution. Wash by filling each well with Wash Buffer (300 μl) using a multi-channel Pipette or microtiter plate washer. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
- 5. Add 100 µl of 1X prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking.
- 6. Discard the solution. Repeat the wash as in step 4.
- 7. Add 100 µl of prepared Streptavidin solution (see Reagent Preparation step 7) to each well. Incubate for 45 minutes at room temperature with gentle shaking.
- 8. Discard the solution. Repeat the wash as in step 4.
- 9. Add 100 µl of TMB One-Step Substrate Reagent (Item H) to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking.
- 10. Add 50 μl of Stop Solution (Item I) to each well. Read at 450 nm immediately.

7. ASSAY PROCEDURE - SUMMARY

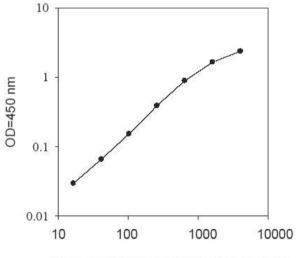
- 1. Prepare all reagents, samples and standards as instructed.
- 2. Add 100 µl standard or sample to each well. Incubate 2.5 hours at room temperature.
- 3. Add 100 µl prepared biotin antibody to each well. Incubate 1 hour at room temperature.
- 4. Add 100 µl prepared Streptavidin solution. Incubate 45 minutes at room temperature.
- 5. Add 100 µl TMB One-Step Substrate Reagent to each well. Incubate 30 minutes at room temperature.
- 6. Add 50 µl Stop Solution to each well. Read at 450 nm immediately.

8. CALCULATIONS

Calculate the mean absorbance for each set of duplicate standards, controls and samples, and subtract the average zero standard optical density. Plot the standard curve on log-log graph paper or using Sigma plot software, with standard concentration on the x-axis and absorbance on the y-axis. Draw the best-fit straight line through the standard points.

8.1 Typical Data

These standard curves are for demonstration only. A standard curve must be run with each assay.



Human uPAR concentration (pg/ml)

8.2 Sensitivity

The minimum detectable dose of Human suPAR was determined to be 15 pg/mL. Minimum detectable dose is defined as the analyte concentration resulting in an absorbance that is 2 standard deviations higher than that of the blank (diluent buffer).

8.3 Spiking & Recovery

Recovery was determined by spiking various levels of Human suPAR into the sample types listed below. Mean recoveries are as follows:

Sample Type	Average % Recovery	Range (%)
Serum	111.2	103-119
Plasma	103.2	90-119
Cell culture media	101.2	90-120

8.4 Linearity

Sample Type		Serum	Plasma	Cell Culture Media	
1:2	Average % of Expected Range (%)	117.5 110-126	113.9 106-122	119.0 111-127	
1:4	Average % of Expected Range (%)	113.3 104-121	118.9 111-127	122.1 114-130	

8.5 Reproducibility

Intra-Assay CV%: <10%

Inter-Assay CV%: <12%

8.6 Specifity

This ELISA kit shows no cross-reactivity with the following cytokines tested: human Angiogenin, BDNF, BLC, CNTF, ENA-78, FGF-4, IL-1 alpha, IL-1 beta, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-11, IL-12 p70, IL-12 p40, IL-13, IL-15, I-309, IP-10, FGF-4, FGF-6, FGF-7, G-CSF, GDNF, GM-CSF, IFN-gamma, IGFBP-2, IGFBP-3, IGFBP-4, Leptin (OB), MCP-1, MCP-2, MCP-3, MDC, MIF, MIG, MIP-1 alpha, MIP-1 beta, MIP-1 delta, PARC, PDGF, RANTES, SCF, SDF-1 alpha, TARC, TGF-alpha, TIMP-1, TIMP-2, TNF-alpha, TNFbeta, TPO, VEGF.

9. TROUBLESHOOTING AND FAQS

Problem	Cause	Solution		
Poor standard curve	Inaccurate pipetting	Check pipettes		
	Improper standard dilution	Briefly centrifuge Item C and dissolve the powder thoroughly by gently mixing		
Low signal	Improper preparation of standard and/or biotinylated antibody	Briefly spin down before opening. Dissolve the powder thoroughly.		
	Too brief incubation times	Ensure sufficient incubation time. Assay procedure step 3 may be done overnight at 4 °C with gentle shaking (note: may increase overall signals including background)		
10	Inadequate reagent volumes or improper dilution	Check pipettes and ensure correct preparation		
Large CV	Inaccurate pipetting	Check pipettes		
	Air bubbles in wells	Remove bubbles in wells		
High background	Plate is insufficiently washed	Review the manual for proper wash. If using a plate washer, ensure that all ports are unobstructed		
	Contaminated wash buffer	Make fresh wash buffer		
Low sensitivity	Improper storage of the ELISA kit	Store your standard at <-70 °C after reconstitution, others at 4 °C. Keep substrate solution protected from light		
	Stop solution	Add stop solution to each well before reading plate		

10. EXPLANATION OF SYMBOLS

REF	Catalogue number
LOT	Batch code
Ţ	Caution
	Use by date
2 °C - 8 °C	Temperature limit
	Manufacturer
www.biovendor.com	Read electronic instructions for use - eIFU
96	The content is sufficient for 96 tests
350 CD	Biological risks

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