

ENG

Instructions for Use:
HUMAN ApoA ELISA

Catalogue number:
REH019R

For research use only!

 **BioVendor**
R&D[®]



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HISTORY OF CHANGES

Previous version	Current version
	ENG.001.A
New edition	

1. INTRODUCTION

The BioVendor ApoA ELISA kit is an in vitro enzyme-linked immunosorbent assay for the quantitative measurement of human ApoA in serum, plasma, and cell culture supernatants. This assay employs an antibody specific for human ApoA coated on a 96-well plate. Standards and samples are pipetted into the wells and ApoA present in a sample is bound to the wells by the immobilized antibody. The wells are washed and biotinylated antihuman ApoA antibody is added. After washing away unbound biotinylated antibody, HRPconjugated streptavidin is pipetted to the wells. The wells are again washed, a TMB substrate solution is added to the wells and color develops in proportion to the amount of ApoA bound. The Stop Solution changes the color from blue to yellow, and the intensity of the color is measured at 450 nm.

2. STORAGE, EXPIRATION

The kit may be stored at 4°C for up to 6 months. For extended storage, it is recommended to store at -80°C. For prepared reagent storage, see table below.

3. REAGENT SUPPLIED

Component	Size / Description	Storage / Stability After Preparation
ApoA Microplate (Item A)	96 wells (12 strips x 8 wells) coated with anti-Human ApoA	1 month at 4 °C*
Wash Buffer Concentrate (20X) (Item B)	25 ml of 20X concentrated solution	1 month at 4 °C
Standard Protein (Item C)	2 vials of Human ApoA. 1 vial is enough to run each standard in duplicate	1 week at -80 °C
Detection Antibody ApoA (Item F)	2 vials of biotinylated anti-Human ApoA. Each vial is enough to assay half the microplate	5 days at 4 °C
HRP-Streptavidin Concentrate (Item G)	200 µl 1000X concentrated HRP-conjugated streptavidin	Do not store and reuse
TMB One-Step Substrate Reagent (Item H)	12 ml of 3,3,5,5'-tetramethylbenzidine (TMB) in buffer solution	N/A
Stop Solution (Item I)	8 ml of 0.2 M sulfuric acid	N/A
Assay Diluent (Item E2)	2 bottles of 15ml of 5X concentrated buffer	1 month at 4 °C

*Return unused wells to the pouch containing desiccant pack, reseal along entire edge.

4. MATERIAL REQUIRED BUT NOT SUPPLIED

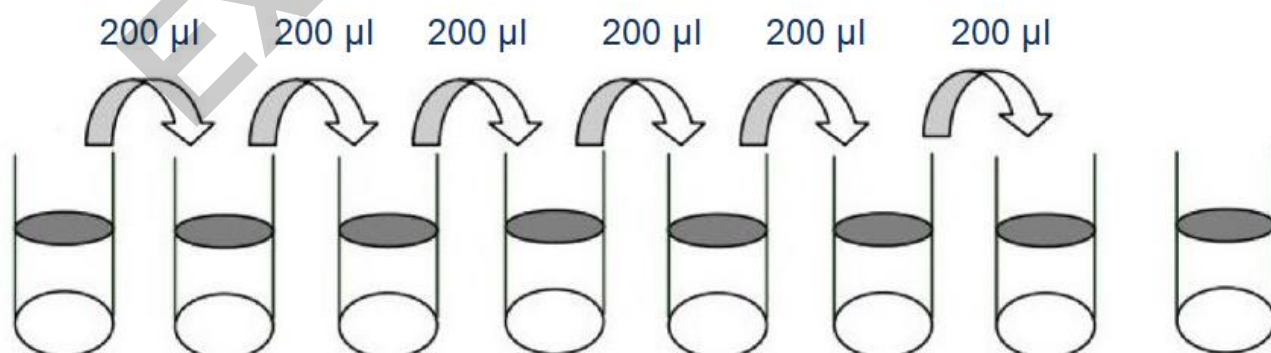
- Microplate reader capable of measuring absorbance at 450 nm.
- Precision pipettes to deliver 2 µl to 1 ml volumes.
- Adjustable 1-25 ml pipettes for reagent preparation.
- 100 ml and 1 liter graduated cylinders.
- Absorbent paper.
- Distilled or deionized water.
- Log-log graph paper or computer and software for ELISA data analysis.
- Tubes to prepare standard or sample dilutions.

5. PREPARATION OF REAGENTS

1. Bring all reagents and samples to room temperature (18 - 25°C) before use.
2. Assay Diluent (Item E2) should be diluted 5-fold with deionized or distilled water before use.
3. Sample dilution: 1X Assay Diluent (Item E2) should be used for dilution of serum, plasma, and cell culture supernatant samples. The suggested dilution for normal serum/plasma is 1,000,000 fold. For example, add 1 µl of serum or plasma into a tube with 999 µl 1X Assay Diluent to prepare a 1,000-fold diluted sample. Mix thoroughly and then pipette 1 µl of prepared 1,000-fold diluted sample into a tube with 999 µl 1X Assay Diluent to prepare a final 1,000,000-fold diluted sample.

Note: Levels of ApoA may vary between different samples. Optimal dilution factors for each sample must be determined by the investigator.

4. Preparation of standard: Briefly spin a vial of Item C. Add 500 µl of 1X Assay Diluent (Item E2, Assay Diluent should be diluted 5-fold with deionized or distilled water before use) into Item C vial to prepare a 20 ng/ml standard. Dissolve the powder thoroughly by a gentle mix. Pipette 300 µl of 1X Assay Diluent into each tube. Use the stock standard solution to produce a dilution series (shown below). Mix each tube thoroughly before the next transfer. 1X Assay Diluent serves as the zero standard (0 ng/ml).



	Std1	Std2	Std3	Std4	Std5	Std6	Std7	Zero Standard
Diluent volume	Item C+ 500 µl	300 µl	300 µ	300 µ	300 µ	300 µ	300 µ	300 µ
Conc.	20 ng/ml	8 ng/ml	3.2 ng/ml	1.28 ng/ml	0.512 ng/ml	0.205 ng/ml	0.082 ng/ml	0 ng/ml

5. If the Wash Concentrate (20X) (Item B) contains visible crystals, warm to room temperature and mix gently until dissolved. Dilute 20 ml of Wash Buffer Concentrate into deionized or distilled water to yield 400 ml of 1X Wash Buffer.
6. Briefly spin the Detection Antibody vial (Item F) before use. Add 100 μ l of 1X Assay Diluent (Item E2) into the vial to prepare a detection antibody concentrate. Pipette up and down to mix gently (the concentrate can be stored at 4°C for 5 days). The detection antibody concentrate should be diluted 80-fold with 1X Assay Diluent (Item E2) and used in step 5 of Part VI Assay Procedure.
7. Briefly spin the HRP-Streptavidin concentrate vial (Item G) and pipette up and down to mix gently before use, as precipitates may form during storage. HRP-Streptavidin concentrate should be diluted 1000-fold with 1X Assay Diluent (Item E2).

For example: Briefly spin the vial (Item G) and pipette up and down to mix gently. Add 10 μ l of HRP-Streptavidin concentrate into a tube with 10 ml 1X Assay Diluent to prepare a 1,000-fold diluted HRP-Streptavidin solution (do not store the diluted solution for next day use). Mix well.

6. ASSAY PROCEDURE

1. Bring all reagents and samples to room temperature (18 - 25°C) before use. It is recommended that all standards and samples be run at least in duplicate.
2. Label removable 8-well strips as appropriate for your experiment.
3. Add 100 μ l of each standard (see Reagent Preparation step 3) and sample into appropriate wells. Cover wells and incubate for 2.5 hours at room temperature with gentle shaking.
4. Discard the solution and wash 4 times with 1X Wash Solution. Wash by filling each well with Wash Buffer (300 μ l) using a multi-channel Pipette or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
5. Add 100 μ l of 1X prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking.
6. Discard the solution. Repeat the wash as in step 4.
7. Add 100 μ l of prepared Streptavidin solution (see Reagent Preparation step 7) to each well. Incubate for 45 minutes at room temperature with gentle shaking.
8. Discard the solution. Repeat the wash as in step 4.
9. Add 100 μ l of TMB One-Step Substrate Reagent (Item H) to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking.
10. Add 50 μ l of Stop Solution (Item I) to each well. Read at 450 nm immediately.

7. ASSAY PROCEDURE - SUMMARY

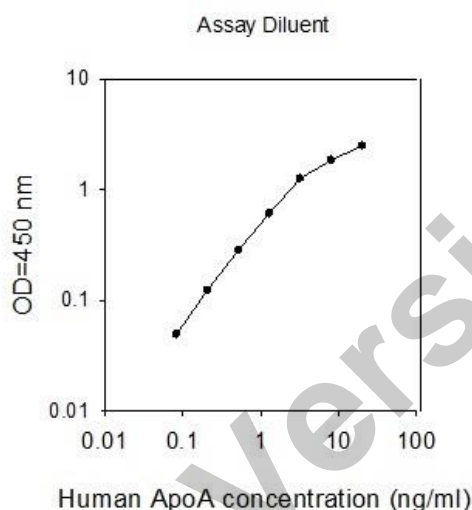
1. Prepare all reagents, samples and standards as instructed.
2. Add 100 μ l standard or sample to each well. Incubate 2.5 hours at room temperature.
3. Add 100 μ l prepared biotin antibody to each well. Incubate 1 hour at room temperature.
4. Add 100 μ l prepared Streptavidin solution. Incubate 45 minutes at room temperature.
5. Add 100 μ l TMB One-Step Substrate Reagent to each well. Incubate 30 minutes at room temperature.
6. Add 50 μ l Stop Solution to each well. Read at 450 nm immediately

8. CALCULATIONS

Calculate the mean absorbance for each set of duplicate standards, controls and samples, and subtract the average zero standard optical density. Plot the standard curve on log-log graph paper or using Sigma plot software, with standard concentration on the x-axis and absorbance on the y-axis. Draw the best-fit straight line through the standard points.

8.1 TYPICAL DATA

These standard curves are for demonstration only. A standard curve must be run with each assay.



8.2 SENSITIVITY

The minimum detectable dose of Human ApoA was determined to be 0.08 ng/ml. Minimum detectable dose is defined as the analyte concentration resulting in an absorbance that is 2 standard deviations higher than that of the blank (diluent buffer).

8.3 SPIKING & RECOVERY

Recovery was determined by spiking various levels of Human ApoA into the sample types listed below. Mean recoveries are as follows:

Sample Type	Average % Recovery	Range (%)
Serum	116.1	93-141
Plasma	134.0	105-141
Cell culture media	141.4	134-144

8.4 LINEARITY

Sample Type	Serum	Plasma	Cell Culture Media
1:2	Average % of Expected Range (%)	111.7 104-119	93.41 89-101
1:4	Average % of Expected Range (%)	99.28 88-110	99.48 75-116
			90.80 78-103
			121.6 116-127

8.5 REPRODUCIBILITY

Intra-Assay CV%: <10%

Inter-Assay CV%: <12%

8.6 SPECIFICITY





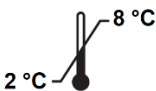




This ELISA antibody pair detects human ApoA1. Cross-reacts with ApoA1 from rhesus and cynomolgus macaques. Other species not determined.

Example Version

9. TROUBLESHOOTING

Problem	Cause	Solution
Poor standard curve	Inaccurate pipetting	Check pipettes
	Improper standard dilution	Briefly centrifuge Item C and dissolve the powder thoroughly by gently mixing
Low signal	Improper preparation of standard and/or biotinylated antibody	Briefly spin down before opening. Dissolve the powder thoroughly.
	Too brief incubation times	Ensure sufficient incubation time. Assay procedure step 3 may be done overnight at 4 °C with gentle shaking (note: may increase overall signals including background)
	Inadequate reagent volumes or improper dilution	Check pipettes and ensure correct preparation
Large CV	Inaccurate pipetting	Check pipettes
	Air bubbles in wells	Remove bubbles in wells
High background	Plate is insufficiently washed	Review the manual for proper wash. If using a plate washer, ensure that all ports are unobstructed
	Contaminated wash buffer	Make fresh wash buffer
Low sensitivity	Improper storage of the ELISA kit	Store your standard at <-70 °C after reconstitution, others at 4 °C. Keep substrate solution protected from light
	Stop solution	Add stop solution to each well before reading plate

10. EXPLANATION OF SYMBOLS

	Catalogue number
	Batch code
	Caution
	Use by date
	Temperature limit
	Manufacturer
 www.biovendor.com	Read electronic instructions for use - eIFU
	The content is sufficient for 96 tests
	Biological risks



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