2

Instructions for Use: Human FGF-23 ELISA

Catalogue number: **RBL007R**

For research use only!





BioVendor - Laboratorní medicína a.s.

Karásek 1767/1, 621 00 Brno, Czech Republic

+420 549 124 185

info@biovendor.com

sales@biovendor.com

www.biovendor.com

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HISTORY OF CHANGES

Previous version	Current version
	ENG.001.A
New edition	

1. INTENDED USE

Enzyme Immunoassay for the quantitative determination of Fibroblast growth factor 23 (FGF23) in human EDTA plasma.

2. STORAGE, EXPIRATION

- The kit must be stored at $2 8^{\circ}$ C.
- The opened components can be stored for one week at 2 8°C

3. INTRODUCTION

Fibroblast growth factor-23 (FGF-23) is a phosphaturic hormone involved in mineral bone metabolism that helps control phosphate homeostasis and reduces 1,25- dihydroxyvitamin D synthesis. Recent data have highlighted the relevant direct FGF-23 effects on the myocardium, and high plasma levels of FGF-23 have been associated with adverse cardiovascular outcomes in humans, such as heart failure and arrhythmias. Therefore, FGF-23 has emerged as a novel biomarker of cardiovascular risk. experimental data suggest FGF-23 as a direct mediator of cardiac hypertrophy development, cardiac fibrosis and cardiac dysfunction via specific myocardial FGF receptor (FGFR) activation[1].

4. TEST PRINCIPLE

The microtiter plate is coated with the antibody specifically binding Fibroblast growth factor 23. The human EDTA plasma is incubated in the plate with the capture antibody.

The specimen is washed out and the specifically bound protein is incubated with biotin-labelled detection antibody. Following another washing step, Streptavidin-HRP conjugate is added into the well

Unbound reagent is then washed out. Horseradish peroxidase (HRP) bound in the complex reacts with the chromogenic substrate (TMB) creating the blue colour. The reaction is stopped by addition of STOP solution (H_2SO_4).

The absorbance values are measured at 450 nm (optionally 450/630 nm) and are proportional to the concentration of FGF23 in the specimen. The concentration of FGF23 in unknown samples is determined from the calibration curve which is created by plotting the absorbance values against the standard concentration values.

5. PRECAUTIONS

- For research use only
- For professional laboratory use
- The reagents with different lot numbers should not be mixed
- To prevent cross sample contamination, use disposable labware and pipette tips

- To protect laboratory stuff, wear protective gloves and protective clothing
- The substrate solution should remain colourless, keep it protected from light
- The test should be performed at standard laboratory conditions (temperature 25°C ± 2°C).

6. REAGENT SUPPLIED

Item	Qty.
Antibody Coated Microtiter Plate	96 wells
Biotin-labelled Antibody	13 ml
Streptavidin-HRP Conjugate	13 ml
Master Standard (lyophilized)	1 vial
Quality Control A (human serum, lyophilized)	1 vial
Quality Control B (human serum, lyophilized)	1 vial
Dilution Buffer	13 ml
Wash Buffer 15x conc.	50 ml
Substrate Solution	13 ml
STOP Solution	13 ml

7. MATERIAL REQUIRED BUT NOT SUPPLIED

- Glassware and test tubes
- Microtiter plate washer
- Precision pipettes (various volumes) with tips
- Orbital shaker
- Microtiter plate reader capable of measuring absorbance at 450 nm or 450/630 nm with software for data generation

8. PREPARATION OF REAGENTS

Use new pipette tip for pipetting different reagents and samples to prevent cross-contamination. All reagents and samples should be allowed to reach the temperature $25 \pm 2^{\circ}$ C.

8.1 Preparation of Standards

Reconstitute lyophilized Human FGF23 Standard in Dilution Buffer, for the volume information see the Certificate of Analysis. Let it rehydrate for 15 min. The concentration of human FGF23 in Master Standard is 2000 pg/ml.

Prepare set of Standard solution as follows:

Use the Master Standard for serial dilution (as below). Mix each tube thoroughly before the next transfer. The Dilution Buffer serves as Blank.

	Volume of Standard	Dilution Buffer	Concentration
Std1	Standard 2000 pg/ml (lyophilized)	See CoA	2000 pg/ml
Std2	300 μL of Std1	300 μL	1000 pg/ml
Std3	180 μL of Std2	450 μL	400 pg/ml
Std4	300 μL of Std3	300 μL	200 pg/ml
Std5	300 μL of Std4	300 µL	100 pg/ml
Std6	300 μL of Std5	300 µL	50 pg/ml
Blank	-	300 μL	0 pg/ml

8.2 Preparation of Quality Control A and B

Reconstitute the lyophilized human serum Quality Controls with deionized/distilled water, for the volume information see the Certificate of Analysis. Let the QCs rehydrate for 15 min and dilute them 1:2 in Dilution Buffer, prior to use, see Preparation of samples.

8.3 Preparation of Wash Buffer 1x

Prepare a working solution of Wash Buffer by adding 50 ml of Wash Buffer 15x conc. to 700 ml of deionized/ distilled water (dH₂O). Mix well. Store at 4°C for two weeks or at -20°C for long term storage.

9. PREPARATION OF SAMPLES

Human EDTA plasma may be used with this assay. For long-term storage the samples should be frozen at minimum -70°C. Lipemic or haemolytic samples may cause false results. Recommended dilution of samples is 1:2, i.e., for singlets 75 μ L of sample + 75 μ L of Dilution Buffer, for duplicates 150 μ L of samples + 150 μ L of Dilution Buffer, respectively. Do not store the diluted samples.

10. ASSAY PROCEDURE

- 1. Prepare the reagents as described in the previous chapter.
- 2. Pipette 100 μL of set of Standards, Quality Controls, diluted Samples and Dilution Buffer = Blank into each well. Incubate for **OVER NIGHT** at 4°C ±2°C, NO shaking.
- 3. Wash the wells 5-times with 1x Wash Buffer (350 µL/well). When finished, tap the plate against the paper towel to remove the liquid completely.
- 4. Pipette 100 μL of Biotin-labelled Antibody into each well. Incubate for **2 hours** at 25°C ±2°C, shaking at **500 rpm**.
- 5. Wash the wells as described in point 3.
- 6. Pipette 100 μL of Streptavidin-HRP into each well. Incubate for **30 min** at 25°C ±2°C, shaking at **300 rpm**.
- 7. Wash the wells as described in point 3.
- 8. Pipette 100 μL Substrate solution, incubate for **15 min**, at 25°C ±2°C. Avoid exposure to the light during this step.
- 9. Pipette 100 µL of STOP solution.
- 10. Read the signal at 450 or 450/630 nm within 15 min.

Plate layout

	1	ı	2	3	4	5	6	7	8	9	10	11	12
Α	S	Sto	11	Bckg	Sa 4	Sa 8	So 12	Sa 16	Sa 20	Sa 24	Sa 28	Sa 32	Sa 36
В	S	Sto	12	БСКУ	Sa 4	Sao	3a 12	Sa 10	Sa 20	Sa 24	Sa 20	Sa 32	Sa 36
C	S	Sto	13	Sa 1	Sa 5	Sa 9	So 12	Sa 17	So 21	So 25	Sa 29	Sa 33	Sa 37
D	S	Sto	14	Sai	Sas	5a 9	Sa 13	Sa 17	Sa 21	Sa 25	Sa 29	Sa 33	3a 31
Ε	S	Sto	15	Sa 2	Sa 6	Sa 10	Sa 14	\$2.19	Sa 22	Sa 26	So 30	Sa 34	Sa 38
F	S	Sto	16	Sa Z	5a 0	Sa 10	Sa 14	Sa 10	3a 22	Sa 20	Sa 30	Sa 34	Sa 30
G	(QC	A	Sa3	Sa 7	Sa 11	Sa 15	Sa 19	Sa 23	Sa 27	Sa 31	Sa 35	Sa 39
Н	(QC	В	Sas	Sa 7	Sall	Sa 15	Sa 19	Sa 23	Sa 21	Sa 31	Sa 35	Sa 39

11. PERFORMANCE CHARACTERISTICS

Samples used in the tests were diluted 1:2 as recommended and assayed. The results are multiplied by the dilution factor.

11.1 Sensitivity

The limit of detection, defined as a concentration of human FGF23 giving absorbance higher than absorbance of blank + 3 standard deviations, is better than 25 pg/ml of sample.

11.2 Precision

11.2.1 Intra-assay

Sample	Mean (pg/ml)	SD	CV (%)
1	1718	100	5.8
2	2 492		3.8

11.2.2 Inter-assay (Run – to – run)

Sample	Mean (pg/ml)	SD	CV (%)
1	1 41		6.3
2	742	90	12.1

11.3 Accuracy

11.3.1 Dilution linearity

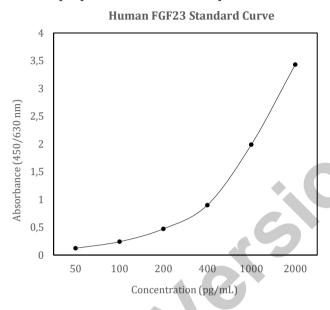
Sample	Dilution	Measured concentration (pg/ml)	Expected concentration (pg/ml)	Yield (%)
		2003	-	-
4	2x	1058	1001	106
l	4x	544	501	109
	8x	284	250	113
		441	<u>-</u>	-
2	2x	231	221	105
2	4x	111	110	101
	8x	61	55	110

11.3.2 Spiking Recovery

Sample	Spike (pg/ml)	Measured concentration (pg/ml)	Expected concentration (pg/ml)	Yield (%)
	-	61	-	-
4	800	749	861	87
	400	421	461	91
	200	225	261	86

12. CALCULATION

The standard curve needs to be measured in every test. Most of the microplate reader can automatically calculate the analyte concentration using 4-parameter algorithm or alternative functions to fit the standard points properly. The concentrations need to be multiplied by the dilution factor, either automatically by reader or manually.



13. REFERENCES

¹ Vázquez-Sánchez S, Poveda J, Navarro-García JA, González-Lafuente L, Rodríguez-Sánchez E, Ruilope LM, Ruiz-Hurtado G. An Overview of FGF-23 as a Novel Candidate Biomarker of Cardiovascular Risk. Front Physiol. 2021 Mar 9;12:632260. doi: 10.3389/fphys.2021.632260. PMID: 33767635; PMCID: PMC7985069.

14. EXPLANATION OF SYMBOLS

REF	Catalogue number
LOT	Batch code
Ţ	Caution
	Use by date
2 °C - 8 °C	Temperature limit
	Manufacturer
www.biovendor.com	Read electronic instructions for use - eIFU
96	The content is sufficient for 96 tests
\$20 P	Biological risks

15. ASSAY PROCEDURE - SUMMARY

Add 100 µL of Standards, diluted QCs and Samples to the wells



Incubate OVER NIGHT at 4°C, NO shaking

3-times wash the wells (350 µL/well)



Add 100 µL of Biotin-labelled Antibody to the wells



Incubate for 2 hours at 25°C, shaking at 500

3-times wash the wells (350 µL/well)



Add 100 µL of SAV-HRP to the wells

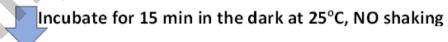


Incubate for 30 min at 25°C, shaking at 300

3-times wash the wells (350 µL/well)



Add 100 µL of Substrate Solution to the wells



Add 100 µL of Stop Solution to the wells



Read the signal at 450 nm (450/630 nm) within 15 min

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+420 549 124 185

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