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Instructions for Use: RAT CRP ELISA

Catalogue number: **REH002R**

For research use only!





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HISTORY OF CHANGES

| Previous version | Current version |
|------------------|-----------------|
| | ENG.001.A |
| New edition | |

1. INTRODUCTION

The BioVendor Rat CRP (C Reactive Protein) ELISA kit is an in vitro enzyme-linked immunosorbent assay for the quantitative measurement of Rat CRP in serum, plasma, cell culture supernatants. This assay employs an antibody specific for Rat CRP coated on a 96-well plate. Standards and samples are pipetted into the wells and CRP present in a sample is bound to the wells by the immobilized antibody. The wells are washed and biotinylated antiRat CRP antibody is added. After washing away unbound biotinylated antibody, HRP-conjugated streptavidin is pipetted to the wells. The wells are again washed, a TMB substrate solution is added to the wells and color develops in proportion to the amount of CRP bound. The Stop Solution changes the color from blue to yellow, and the intensity of the color is measured at 450 nm.

2. STORAGE, EXPIRATION

The kit may be stored at 4°C for up to 6 months. For extended storage, it is recommended to store at -80°C. For prepared reagent storage, see table below.

3. REAGENT SUPPLIED

| Component | Size / Description | Storage / Stability After Preparation | | | | |
|--|--|---------------------------------------|--|--|--|--|
| CRP Microplate (Item A) | 96 wells (12 strips x 8 wells) coated with anti-Rat CRP | 1 month at 4 °C | | | | |
| Wash Buffer Concentrate (20X) (Item B) | 25 ml of 20X concentrated solution | 1 month at 4 °C | | | | |
| Standard Protein (Item C) | 2 vials of Rat CRP. 1 vial is enough to run each standard in duplicate | 1 week at -80 °C | | | | |
| Detection Antibody CRP (Item F) | 2 vials of biotinylated anti-Rat CRP. Each vial is enough to assay half the microplate | 5 days at 4 °C | | | | |
| HRP-Streptavidin Concentrate (Item G) | 200 µl 500X concentrated HRP-conjugated strepavidin | Do not store and reuse | | | | |
| TMB One-Step Substrate Reagent (Item H) | 12 ml of 3,3,5,5'-tetramethylbenzidine (TMB) in buffer solution | N/A | | | | |
| Stop Solution (Item I) | 8 ml of 0.2 M sulfuric acid | N/A | | | | |
| Assay Diluent (Item E2) | 2 bottles of 15ml 5X concentrated buffer | 1 month at 4 °C | | | | |

Note: Return unused wells to the pouch containing desiccant pack, reseal along entire edge.

4. MATERIAL REQUIRED BUT NOT SUPPLIED

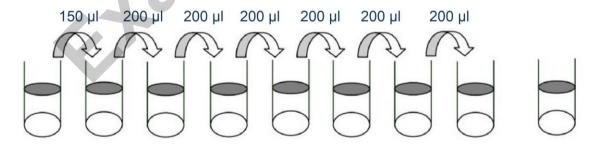
- 1. Microplate reader capable of measuring absorbance at 450 nm.
- 2. Precision pipettes to deliver 2 µl to 1 ml volumes.
- 3. Adjustable 1-25 ml pipettes for reagent preparation.
- 4. 100 ml and 1 liter graduated cylinders.
- 5. Absorbent paper.
- 6. Distilled or deionized water.
- 7. Log-log graph paper or computer and software for ELISA data analysis.
- 8. Tubes to prepare standard or sample dilutions.

5. PREPARATION OF REAGENTS

- 1. Bring all reagents and samples to room temperature (18 25°C) before use.
- 2. Assay Diluent (Item E2) should be diluted 5-fold with deionized or distilled water before use.
- 3. Sample dilution: 1X Assay Diluent (Item E2) should be used for dilution of serum, plasma, and cell culture supernatant samples. The suggested dilution for normal serum/plasma is 20,000 200,000 fold. For example, add 1 µl of serum/plasma into a tube with 99 µl 1X Assay Diluent to prepare a 100-fold diluted sample. Mix thoroughly and then pipette 1 µl of prepared 100-fold diluted sample into a tube with 499 µl 1X Assay Diluent to prepare a final 50,000 fold diluted sample.

<u>Note:</u> Levels of CRP may vary between different samples. Optimal dilution factors for each sample must be determined by the investigator.

4. Preparation of standard: Briefly spin a vial of Item C. Add 400 μI 1X Assay Diluent (Item E2) into Item C vial to prepare a 200 ng/ml standard solution. Dissolve the powder thoroughly by a gentle mix. Add 150 μI CRP standard from the vial of tem C, into a tube with 350 μI 1X Assay Diluent to prepare a 60 ng/ml standard solution. Pipette 300 μI 1X Assay Diluent into each tube. Use the 60 ng/ml standard solution to produce a dilution series (shown below). Mix each tube thoroughly before the next transfer. 1X Assay Diluent serves as the zero standard (0 pg/ml).



| | | Std1 | Std2 | Std3 | Std4 | Std5 | Std6 | Std7 | Zero Standard |
|----------------|-------------------|-------------|-------------|--------------|---------------|----------------|----------------|----------------|------------------|
| Diluent volume | Item C +400 µl | 350 µl | 300 µl | 300 µl | 300 µl | 300 µl | 300 µl | 300 µl | 300 µl |
| Conc. | 200 ng/ml | 60 ng/ml | 24 ng/ml | 9.6 ng/ml | 3.84 ng/ml | 1.536 ng/ml | 0.614 ng/ml | 0.246 ng/ml | 0 ng/ml |

- 5. If the Wash Concentrate (20X) (Item B) contains visible crystals, warm to room temperature and mix gently until dissolved. Dilute 20 ml of Wash Buffer Concentrate into deionized or distilled water to yield 400 ml of 1X Wash Buffer.
- 6. Briefly spin the Detection Antibody vial (Item F) before use. Add 100 µl of 1X Assay Diluent (Item E2) into the vial to prepare a detection antibody concentrate. Pipette up and down to

- mix gently (the concentrate can be stored at 4°C for 5 days). The detection antibody concentrate should be diluted 80-fold with 1X Assay Diluent (Item E2) and used in step 5 of Part VI Assay Procedure.
- 7. Briefly spin the HRP-Streptavidin concentrate vial (Item G) and pipette up and down to mix gently before use, as precipitates may form during storage. HRP-Streptavidin concentrate should be diluted 500-fold with 1X Assay Diluent (Item E2). For example: Briefly spin the vial (Item G) and pipette up and down to mix gently. Add 20 µl of HRP-Streptavidin concentrate into a tube with 10 ml 1X Assay Diluent to prepare a 500-fold diluted HRP-Streptavidin solution (don't store the diluted solution for next day

6. ASSAY PROCEDURE

use). Mix well.

- 1. Bring all reagents and samples to room temperature (18 25°C) before use. It is recommended that all standards and samples be run at least in duplicate.
- 2. Label removable 8-well strips as appropriate for your experiment.
- 3. Add 100 µl of each standard (see Reagent Preparation step 3) and sample into appropriate wells. Cover wells and incubate for 2.5 hours at room temperature with gentle shaking.
- 4. Discard the solution and wash 4 times with 1X Wash Solution. Wash by filling each well with Wash Buffer (300 µl) using a multi-channel Pipette or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
- 5. Add 100 µl of 1X prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking.
- 6. Discard the solution. Repeat the wash as in step 4.
- 7. Add 100 µl of prepared Streptavidin solution (see Reagent Preparation step 7) to each well. Incubate for 45 minutes at room temperature with gentle shaking.
- 8. Discard the solution. Repeat the wash as in step 4.
- 9. Add 100 µl of TMB One-Step Substrate Reagent (Item H) to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking.
- 10. Add 50 µl of Stop Solution (Item I) to each well. Read at 450 nm immediately.

7. ASSAY PROCEDURE - SUMMARY

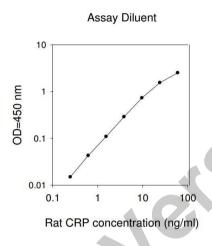
- 1. Prepare all reagents, samples and standards as instructed.
- 2. Add 100 µl standard or sample to each well. Incubate 2.5 hours at room temperature.
- 3. Add 100 µl prepared biotin antibody to each well. Incubate 1 hour at room temperature.
- 4. Add 100 µl prepared Streptavidin solution. Incubate 45 minutes at room temperature.
- 5. Add 100 µl TMB One-Step Substrate Reagent to each well. Incubate 30 minutes at room temperature.
- 6. Add 50 µl Stop Solution to each well. Read at 450 nm immediately.

8. CALCULATIONS

Calculate the mean absorbance for each set of duplicate standards, controls and samples, and subtract the average zero standard optical density. Plot the standard curve on log-log graph paper or using Sigma plot software, with standard concentration on the x-axis and absorbance on the y-axis. Draw the best-fit straight line through the standard points.

8.1 Typical Data

These standard curves are for demonstration only. A standard curve must be run with each assay.



8.2 Sensitivity

The minimum detectable dose of Rat CRP was determined to be 200 pg/ml.

Minimum detectable dose is defined as the analyte concentration resulting in an absorbance that is 2 standard deviations higher than that of the blank (diluent buffer).

8.3 Spiking & Recovery

Recovery was determined by spiking various levels of Rat CRP into the sample types listed below. Mean recoveries are as follows:

| Sample Type | Average % Recovery | Range (%) | | |
|--------------------|--------------------|-----------|--|--|
| Serum | 98.54 | 86-108 | | |
| Plasma | 121.5 | 109-142 | | |
| Cell culture media | 112.2 | 104-120 | | |

8.4 Linearity

| Sam | ple Type | Serum | Plasma | Cell Culture Media |
|-----|---------------------------------|----------------|-----------------|-----------------------|
| 1:2 | Average % of Expected Range (%) | 82.16 70-94 | 101.7 93-120 | 101.1 93-109 |
| 1:4 | Average % of Expected Range (%) | 86.50 80-92 | 75.42 67-83 | 83.48 68-90 |

8.5 Reproducibility

Intra-Assay CV%: <10%

Inter-Assay CV%: <12%

8.6 Specifity

This ELISA kit shows no cross-reactivity with the following cytokines tested: rat CINC-2, CINC-3, CNTF, Fractalkine, IL-1 alpha, IL-1 beta, IL-4, IL-6, IL-10, GM-CSF, IFN-gamma, Leptin, Lix, MCP-1, MIP-3 alpha, beta- NGF, TIMP-1, TNF-alpha

9. TROUBLESHOOTING

| Problem | Cause | Solution | | |
|---------------------|---|---|--|--|
| | Inaccurate pipetting | Check pipettes | | |
| Poor standard curve | Improper standard dilution | Briefly centrifuge Item C and dissolve the powder thoroughly by gently mixing | | |
| | Improper preparation of standard and/or biotinylated antibody | Briefly spin down before opening. Dissolve the powder thoroughly. | | |
| Low signal | Too brief incubation times | Ensure sufficient incubation time. Assay procedure step 3 may be done overnight at 4 °C with gentle shaking (note: may increase overall signals including background) | | |
| | Inadequate reagent volumes | Check pipettes and ensure | | |
| | or improper dilution | correct preparation | | |
| Large CV | Inaccurate pipetting | Check pipettes | | |
| J | Air bubbles in wells | Remove bubbles in wells | | |
| High background | Plate is insufficiently washed | Review the manual for proper wash. If using a plate washer ensure that all ports are unobstructed | | |
| | Contaminated wash buffer | Make fresh wash buffer | | |
| Low sensitivity | Improper storage of the ELISA kit | Store your standard at <-70 °C after reconstitution, others at 4 °C. Keep substrate solution protected from light | | |
| | Stop solution | Add stop solution to each well before reading plate | | |

10. EXPLANATION OF SYMBOLS

| REF | Catalogue number | | | | |
|-------------------|---|--|--|--|--|
| LOT | Batch code | | | | |
| Ţ | Caution | | | | |
| | Use by date | | | | |
| 2 °C - 8 °C | Temperature limit | | | | |
| | Manufacturer | | | | |
| www.biovendor.com | Read electronic instructions for use - eIFU | | | | |
| 96 | The content is sufficient for 96 tests | | | | |
| \$20 P | Biological risks | | | | |

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